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Force Field Parametrization of the $\text{Fe}_2+4\text{S}_2-4$ Clusters in Dihydropyrimidine Dehydrogenase, the 5-Fluorouracil Cancer Drug Metabolizer: Facilitating In Silico Pharmacogenomic Studies

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Dihydropyrimidine dehydrogenase (DPD), a dimeric enzyme has attracted significant scientific attention for its role in metabolizing 5 fluorouracil (5-FU) anticancer drug in humans. These metalloprotein consist of highly specialized $4 \times \text{Fe}_2+4\text{S}_2-4$ clusters per monomer. A finely orchestrated inter- and intra-monomer electron transfer chain reaction through these clusters activates a redox metabolic reaction of the rate-limiting step in pyrimidine degradation. During this process, the majority of administered 5-FU drug is catabolically deactivated, while the remaining amount is activated to nucleic acid antimetabolites. DPD mutations in the general population, including people of African descent causes adverse toxicity effects in patients due to altered 5-FU metabolism. Therefore, understanding the impact of mutation on the catalytic mechanism of DPD protein is crucial, particularly in precision oncology medicine. Molecular dynamics simulations, principal component analysis, and dynamic residue network analysis can be used to decipher the effects of missense mutations in proteins. However, the $\text{Fe}_2+4\text{S}_2-4$ clusters in DPD protein require additional force field parameters to adequately describe the Fe_2+ center coordination geometry. Consequently, new AMBER force field parameters for DPD enzyme Fe_2+ center were derived using the original Seminario method and collation features visual force field derivation toolkit (VFFDT) as a supportive approach. The overall study took ~ 2400 CPU hours and was run on 240 cores at the Centre for High-Performance Computing (CHPC) in Cape Town, South Africa. Validation of the derived parameters with a 150 ns MD simulation revealed that both methods produced force field parameters that accurately described the $\text{Fe}_2+4\text{S}_2-4$ cluster architecture of the human DPD protein. These findings lay the groundwork not only for future research in the fields of in silico cancer pharmacogenomics studies, but also for novel drug discovery and repurposing studies involving the human DPD enzyme.

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